# Childhood Immune Maturation and Allergy Development: Regulation by Maternal Immunity and Microbial Exposure

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# Childhood immune maturation and allergy development: Regulation by maternal immunity and microbial exposure

Running head: Maternal immunity and childhood allergy

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#### **Abstract**

The increasing allergy prevalence in affluent countries may be caused by reduced microbial stimulation, resulting in an abnormal postnatal immune maturation. Most studies investigating the underlying mechanisms have focused on postnatal microbial exposure. Also the maternal microbial environment during pregnancy may program the immune development of the child, however. Prenatal environmental exposures may alter gene expression via epigenetic mechanisms, aiming to induce physiological adaptations to the anticipated postnatal environment, but potentially also increasing disease susceptibility in the offspring. Although the importance of fetal programming mostly has been studied in cardiovascular and metabolic disease, this hypothesis is also very attractive in the context of environmentally influenced immune-mediated diseases. This review focuses on how maternal immunity and microbial exposures regulate childhood immune and allergy development. Efficacious preventive measures, required to combat the allergy epidemic, may be identified by determining how the immune interaction between mother and child is influenced by microbial factors.

# **Key words**

Fetal programming, allergy, immune regulation, epigenetics, microbial exposure

# Introduction

Allergic diseases have become a major public health problem in affluent societies <sup>1,2</sup>. Asthma is the most common chronic disease among children, with a major impact on both the physiological and psychological well-being of young children <sup>3</sup>, as well as on socio-economic costs due to hospital admittance, treatment costs and parental sick leave <sup>4</sup>. The allergy epidemic must be counteracted by research identifying successful preventive measures, which do not exist today.

#### The allergic march

Allergic diseases are characterized by inappropriate immune responses to innocuous foreign proteins, allergens. Atopy is defined as personal and/or familiar tendency to produce IgE antibodies to allergens, *i e* become sensitized <sup>5</sup>. The excessive Th2-like responses to allergens in atopic individuals include high production of IgE-inducing IL-4 and IL-13 and eosinophilia-enhancing IL-5 and IL-9 <sup>6,7</sup>. During the early phase of the IgE-mediated allergic reaction, allergen crosslinking of IgE antibodies on mast cells and basophils triggers release of inflammatory mediators <sup>7</sup>. Cytotoxic mediators from eosinophils are important in the late phase reaction, and lead to chronic inflammation <sup>7</sup>.

Atopic eczema, bronchial asthma, allergic rhinoconjunctivitis and immediate types of urticaria and food allergy all belong to the allergic diseases. The allergic march typically begins with the development of IgE antibodies to food allergens accompanied with symptoms of atopic eczema and food allergy <sup>8</sup>. After sensitization during infancy, most children develop tolerance to food allergens <sup>8</sup>. Later in childhood, inhalant allergen sensitization develops together with asthmatic symptoms, while onset of allergic rhinoconjunctivitis is usually seen from early school age <sup>8</sup>.

## Reduced microbial stimulation and the allergy epidemic

As changes in the genotype cannot explain the rapid increase in the allergy prevalence, loss of protective factors or appearance of risk factors in the environment may contribute to the increased prevalence of these diseases since the middle of the last century. A reduced microbial pressure, resulting in insufficient induction of T cells with regulatory and/or Th1-like properties to counteract allergy-inducing Th2 response, may underlie the allergy epidemic <sup>9-13</sup>. Most studies investigating the underlying mechanisms have focused on postnatal microbial exposure <sup>14-18</sup>.

An increasing body of evidence from studies of others and us suggests that the maternal microbial environment during pregnancy can program the immune development of the child, however <sup>13, 19, 20</sup>. Thus, experimental murine models demonstrate that maternal treatment with lipopolysaccharide <sup>21-23</sup> or the commensal *Acinetobacter lwoffii* <sup>24</sup> during gestation attenuates allergic sensitization and airway inflammation in the offspring. Also, epidemiological studies indicate that maternal farm environment exposure during pregnancy protects against allergic sensitization and disease, whereas exposures during infancy alone have weaker or no effect at all <sup>13, 25, 26</sup>. Continued enhanced postnatal microbial exposure may be required for optimal allergy protection, however <sup>26</sup>. Furthermore, in human allergy intervention studies, probiotic supplementation to the mother during pregnancy, as well as to her baby postnatally, may be important for preventive effects <sup>27, 28</sup>. Thus, a preventive effect on atopic eczema has primarily been demonstrated in studies by us and others where probiotics were given both pre- and postnatally <sup>19, 29-33</sup>, whereas two studies with postnatal supplementation only failed to prevent allergic disease 34, 35. Prenatal probiotic supplementation was not given until 36 weeks of gestation in any of the studies, however <sup>19, 29-33</sup>. If prenatal microbial exposure is vital for the preventive effect, starting supplementation already from the second

trimester of pregnancy, when circulating fetal T cells have developed <sup>36</sup>, may have a more powerful preventive effect on allergy development.

### **Epigenetic regulation**

Regulation by epigenetic mechanisms, heritable changes in gene expression occurring without alterations in the DNA sequences <sup>37</sup>, a kind of cellular memory, may play a major role in prenatal immune programming <sup>38</sup>. Epigenetic modifications determine the degree of DNA compaction and accessibility for gene transcription, thus resulting in changes in gene expression that are subsequently passed to somatic daughter cells during mitosis <sup>37</sup>. The main processes modulating DNA accessibility to establish epigenetic memory occur via posttranslational histone modifications and methylation of DNA CpG dinucleotides <sup>37</sup>. DNA methylation, associated with transcriptional repression, is more rigid than histone modifications, with DNA methyltransferases conferring covalent methyl modifications to evolutionary conserved regulatory gene elements, CpG islands <sup>39</sup>. The methylation pattern is thus preserved with high fidelity through cell divisions, assuring preservation of cellular inheritance <sup>39</sup>.

#### **Epigenetic regulation of childhood immune development**

Prenatal environmental exposures may alter gene expression via epigenetic mechanisms, aiming to induce physiological adaptations to the anticipated postnatal environment, but potentially also increasing disease susceptibility in the offspring <sup>40</sup>. This "Developmental Origins of Health and Disease" hypothesis <sup>40</sup> was originally proposed by David Barker <sup>41</sup>. Although the importance of fetal programming mostly has been studied in cardiovascular and metabolic disease <sup>40</sup>, this hypothesis is also very attractive in the context of environmentally influenced immune-mediated diseases. The maternal microbial environment during pregnancy may program the immune development of the child <sup>20</sup>, via epigenetic mechanisms, regulating appropriate maturation of innate immunity <sup>24</sup>,

 $^{25}$  and T helper and regulatory responses  $^{12,42}$ . Th1, Th2 and Th17 differentiation is under epigenetic control  $^{43-45}$ , and human T regulatory cell commitment requires demethylation of the *FOXP3* promoter  $^{46}$ .

# The role of maternal microbial exposure and immune regulation in childhood allergy development

Epigenetically regulated childhood immune development by maternal microbial exposure is likely induced via changes in maternal immune regulation <sup>22, 24</sup>, as there is a close immunological interaction between the mother and her offspring during pregnancy 47, 48. The placenta allows a cross-talk between maternal stimuli, possibly induced via microbial stimulation of maternal Toll-like receptors, and fetal responses <sup>24</sup>. As fetal T cells have developed during the second trimester of gestation <sup>36</sup>, maternal signals may then direct the immune cell lineage commitment of the offspring during a critical developmental period when the epigenetic program is highly susceptible to environmental influences <sup>20</sup>. During pregnancy, the fetal-maternal interface is characterized by high levels of Th2-like cytokines 49 and enrichment of T regulatory cells 50, most likely functioning to divert the maternal immune response away from damaging Th1mediated immunity 51. The association of cord blood IgE levels and neonatal IFN-γ production with maternal but not paternal atopic heredity <sup>52, 53</sup> may depend on an even stronger Th2-deviation in atopic than non-atopic pregnant women <sup>54, 55</sup>. As the cytokine milieu shapes the T helper differentiation, particularly during naïve as compared to established responses <sup>56</sup>, the neonatal immune system is Th2-skewed <sup>57</sup>. The Th2 cytokine locus of in murine neonatal CD4+ T cells is poised epigenetically for rapid and robust production of IL-4 and IL-13 <sup>58</sup>. We have shown an even more marked neonatal Th2-skewing in infants later developing allergic disease 48, possibly due to prenatal epigenetic effects via maternal immune regulation that may be possible to redress by enhanced microbial exposure, *e g* via probiotic supplementation, during pregnancy. The Th2-bias of the new-born should then develop toward a more balanced immune phenotype, including maturation of Th1-like responses <sup>12</sup> and appropriate development of regulatory T cell responses <sup>11</sup>. In farm studies, contact with multiple animal species during pregnancy is positively correlated to Treg cell function and IFN-γ production at birth and with innate immune receptor expression at birth and during childhood <sup>13, 25, 42, 59, 60</sup>. A failure of Th2-silencing during maturation of the immune system may underlie development of Th2-mediated allergic disease <sup>61</sup>. Appropriate microbial stimulation, both pre- and postnatally, may be required to avoid this pathophysiological process <sup>26</sup>.

In this respect, the gut microbiota is quantitatively the most important source of microbial stimulation and may provide a primary signal for the maturation of a balanced postnatal innate and adaptive immune system 62,63. It is likely that our immune system has evolved as much to manage and exploit beneficial microbes as to fend off pathogens <sup>64,65</sup>. The gut microbiota differs during the first months of life in children who later do or do not develop allergic disease 66-68, and the diversity of the microbiota may play an important role in regulating allergy 69,70 and mucosal immune development <sup>63</sup>. To what extent the maternal gut microbiota composition influences that of her offspring is not yet fully clear. Differences in microbiota composition depending on delivery mode do indicate a mother-child transmission of microbiota during vaginal delivery <sup>71,72</sup>. Due to the vast complexity of the gut microbiota, more detailed, basic microbial ecology studies, now made possible by advances in DNA sequencing technologies <sup>73, 74</sup>, in clinically and immunologically well-characterized children and their mothers are needed, however. Also, how the maternal gut microbiota impacts the development of the microbiota of the child, in addition to the effects on immune maturation during infancy, needs further investigation.

#### Conclusion

The maternal microbial environment during pregnancy may program the immune development of the child. Prenatal environmental exposures may alter gene expression via epigenetic mechanisms, aiming to induce physiological adaptations to the anticipated postnatal environment, but potentially also increasing disease susceptibility in the offspring. Efficacious preventive measures, required to combat the allergy epidemic, may be identified by determining how the immune interaction between mother and child is influenced by microbial factors.

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#### References

- Asher MI, Montefort S, Björkstén B, Lai CK, Strachan DP, Weiland SK, Williams H: Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet* 2006;368:733-743.
- Eder W, Ege MJ, von Mutius E: The asthma epidemic. *N Engl J Med* 2006;**355**:2226-2235.
- Rydström I, Dalheim-Englund AC, Holritz-Rasmussen B, Möller C, Sandman PO: Asthma--quality of life for Swedish children. *J Clin Nurs* 2005;**14**:739-749.
- Jansson SA, Arnlind MH, Dahlén SE, Lundbäck B: [Costs of asthma and allergies to society unknown. Cost studies can give better planning of health care and research]. *Läkartidningen* 2007;**104**:2792-2796.
- Johansson SG, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF, Motala C, Ortega Martell JA, Platts-Mills TA, Ring J, Thien F, Van Cauwenberge P, Williams HC: Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. J Allergy Clin Immunol 2004;113:832-836.
- Jenmalm MC, Van Snick J, Cormont F, Salman B: Allergen-induced Th1 and Th2 cytokine secretion in relation to specific allergen sensitization and atopic symptoms in children. *Clin Exp Allergy* 2001;**31**:1528-1535.

- 7 Kim HY, DeKruyff RH, Umetsu DT: The many paths to asthma: phenotype shaped by innate and adaptive immunity. *Nat Immunol* 2010;**11**:577-584.
- Hattevig G, Kjellman B, Björkstén B: Appearance of IgE antibodies to ingested and inhaled allergens during the first 12 years of life in atopic and non-atopic children. *Pediatr Allergy Immunol* 1993;4:182-186.
- 9 Schaub B, Lauener R, von Mutius E: The many faces of the hygiene hypothesis. *J Allergy Clin Immunol* 2006;**117**:969-977.
- Böttcher MF, Jenmalm MC, Voor T, Julge K, Holt PG, Björkstén B: Cytokine responses to allergens during the first 2 years of life in Estonian and Swedish children. *Clin Exp Allergy* 2006;**36**:619-628.
- 11 Lloyd CM, Hawrylowicz CM: Regulatory T cells in asthma. *Immunity* 2009;**31**:438-449.
- Vuillermin PJ, Ponsonby AL, Saffery R, Tang ML, Ellis JA, Sly P, Holt P: Microbial exposure, interferon gamma gene demethylation in naive T-cells, and the risk of allergic disease. *Allergy* 2009;**64**:348-353.
- von Mutius E, Vercelli D: Farm living: effects on childhood asthma and allergy. *Nat Rev Immunol* 2010;**10**:861-868.
- Böttcher MF, Björkstén B, Gustafson S, Voor T, Jenmalm MC: Endotoxin levels in Estonian and Swedish house dust and atopy in infancy. *Clin Exp Allergy* 2003;**33**:295-300.
- Bashir ME, Louie S, Shi HN, Nagler-Anderson C: Toll-like receptor 4 signaling by intestinal microbes influences susceptibility to food allergy. *J Immunol* 2004;**172**:6978-6987.
- Foliaki S, Pearce N, Björkstén B, Mallol J, Montefort S, von Mutius E: Antibiotic use in infancy and symptoms of asthma, rhinoconjunctivitis, and eczema in children 6 and 7 years old: International Study of Asthma and Allergies in Childhood Phase III. *J Allergy Clin Immunol* 2009;**124**:982-989.
- 17 Kwon HK, Lee CG, So JS, Chae CS, Hwang JS, Sahoo A, Nam JH, Rhee JH, Hwang KC, Im SH: Generation of regulatory dendritic cells and CD4+Foxp3+ T cells by probiotics administration suppresses immune disorders. *Proc Natl Acad Sci U S A* 2010;**107**:2159-2164.
- Ege MJ, Mayer M, Normand AC, Genuneit J, Cookson WO, Braun-Fahrlander C, Heederik D, Piarroux R, von Mutius E: Exposure to environmental microorganisms and childhood asthma. *N Engl J Med* 2011;**364**:701-709.
- Abrahamsson TR, Jakobsson T, Böttcher MF, Fredrikson M, Jenmalm MC, Björkstén B, Oldaeus G: Probiotics in prevention of IgE-associated eczema: a double-blind, randomized, placebo-controlled trial. *J Allergy Clin Immunol* 2007;**119**:1174-1180.
- Hawrylowicz C, Ryanna K: Asthma and allergy: the early beginnings. *Nat Med* 2010;**16**:274-275.
- Blümer N, Herz U, Wegmann M, Renz H: Prenatal lipopolysaccharide-exposure prevents allergic sensitization and airway inflammation, but not airway responsiveness in a murine model of experimental asthma. *Clin Exp Allergy* 2005;**35**:397-402.
- Gerhold K, Avagyan A, Seib C, Frei R, Steinle J, Ahrens B, Dittrich AM, Blumchen K, Lauener R, Hamelmann E: Prenatal initiation of endotoxin airway

- exposure prevents subsequent allergen-induced sensitization and airway inflammation in mice. *J Allergy Clin Immunol* 2006;**118**:666-673.
- Cao L, Wang J, Zhu Y, Tseu I, Post M: Maternal endotoxin exposure attenuates allergic airway disease in infant rats. *Am J Physiol Lung Cell Mol Physiol* 2011;**In press**.
- 24 Conrad ML, Ferstl R, Teich R, Brand S, Blumer N, Yildirim AO, Patrascan CC, Hanuszkiewicz A, Akira S, Wagner H, Holst O, von Mutius E, Pfefferle PI, Kirschning CJ, Garn H, Renz H: Maternal TLR signaling is required for prenatal asthma protection by the nonpathogenic microbe *Acinetobacter lwoffii* F78. *J Exp Med* 2009;**206**:2869-2877.
- Ege MJ, Bieli C, Frei R, van Strien RT, Riedler J, Ublagger E, Schram-Bijkerk D, Brunekreef B, van Hage M, Scheynius A, Pershagen G, Benz MR, Lauener R, von Mutius E, Braun-Fahrlander C: Prenatal farm exposure is related to the expression of receptors of the innate immunity and to atopic sensitization in school-age children. *J Allergy Clin Immunol* 2006;**117**:817-823.
- Douwes J, Cheng S, Travier N, Cohet C, Niesink A, McKenzie J, Cunningham C, Le Gros G, von Mutius E, Pearce N: Farm exposure in utero may protect against asthma, hay fever and eczema. *Eur Respir J* 2008;**32**:603-611.
- Lee J, Seto D, Bielory L: Meta-analysis of clinical trials of probiotics for prevention and treatment of pediatric atopic dermatitis. *J Allergy Clin Immunol* 2008;**121**:116-121 e111.
- Tang ML, Lahtinen SJ, Boyle RJ: Probiotics and prebiotics: clinical effects in allergic disease. *Curr Opin Pediatr* 2010;**22**:626-634.
- 29 Kalliomäki M, Salminen S, Arvilommi H, Kero P, Koskinen P, Isolauri E: Probiotics in primary prevention of atopic disease: a randomised placebocontrolled trial. *Lancet* 2001;**357**:1076-1079.
- 30 Kalliomäki M, Salminen S, Poussa T, Arvilommi H, Isolauri E: Probiotics and prevention of atopic disease: 4-year follow-up of a randomised placebocontrolled trial. *Lancet* 2003;**361**:1869-1871.
- 31 Kukkonen K, Savilahti E, Haahtela T, Juntunen-Backman K, Korpela R, Poussa T, Tuure T, Kuitunen M: Probiotics and prebiotic galacto-oligosaccharides in the prevention of allergic diseases: a randomized, double-blind, placebo-controlled trial. *J Allergy Clin Immunol* 2007;**119**:192-198.
- Wickens K, Black PN, Stanley TV, Mitchell E, Fitzharris P, Tannock GW, Purdie G, Crane J: A differential effect of 2 probiotics in the prevention of eczema and atopy: a double-blind, randomized, placebo-controlled trial. *J Allergy Clin Immunol* 2008;**122**:788-794.
- 33 Kim JY, Kwon JH, Ahn SH, Lee SI, Han YS, Choi YO, Lee SY, Ahn KM, Ji GE: Effect of probiotic mix (Bifidobacterium bifidum, Bifidobacterium lactis, Lactobacillus acidophilus) in the primary prevention of eczema: a double-blind, randomized, placebo-controlled trial. *Pediatr Allergy Immunol* 2010;**21**: e386–e393.
- Taylor AL, Dunstan JA, Prescott SL: Probiotic supplementation for the first 6 months of life fails to reduce the risk of atopic dermatitis and increases the risk of allergen sensitization in high-risk children: a randomized controlled trial. *J Allergy Clin Immunol* 2007;**119**:184-191.
- Soh SE, Aw M, Gerez I, Chong YS, Rauff M, Ng YP, Wong HB, Pai N, Lee BW, Shek LP: Probiotic supplementation in the first 6 months of life in at risk Asian

- infants--effects on eczema and atopic sensitization at the age of 1 year. *Clin Exp Allergy* 2009;**39**:571-578.
- Papiernik M: Correlation of lymphocyte transformation and morphology in the human fetal thymus. *Blood* 1970;**36**:470-479.
- Jaenisch R, Bird A: Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nat Genet* 2003;**33 Suppl**:245-254.
- Martino D, Prescott S: Epigenetics and prenatal influences on asthma and allergic airways disease. *Chest* 2011;**139**:640-647.
- 39 Kim JK, Samaranayake M, Pradhan S: Epigenetic mechanisms in mammals. *Cell Mol Life Sci* 2009;**66**:596-612.
- Wadhwa PD, Buss C, Entringer S, Swanson JM: Developmental origins of health and disease: brief history of the approach and current focus on epigenetic mechanisms. *Semin Reprod Med* 2009;**27**:358-368.
- Barker DJ: The fetal and infant origins of adult disease. *BMJ* 1990;**301**:1111.
- 42 Schaub B, Liu J, Hoppler S, Schleich I, Huehn J, Olek S, Wieczorek G, Illi S, von Mutius E: Maternal farm exposure modulates neonatal immune mechanisms through regulatory T cells. *J Allergy Clin Immunol* 2009;**123**:774-782.
- Wilson CB, Rowell E, Sekimata M: Epigenetic control of T-helper-cell differentiation. *Nat Rev Immunol* 2009;9:91-105.
- Janson PC, Winerdal ME, Winqvist O: At the crossroads of T helper lineage commitment Epigenetics points the way. *Biochim Biophys Acta* 2009;**1790**:906-919.
- Janson PC, Linton LB, Bergman EA, Marits P, Eberhardson M, Piehl F, Malmström V, Winqvist O: Profiling of CD4+ T cells with epigenetic immune lineage analysis. *J Immunol* 2011;**186**:92-102.
- Janson PC, Winerdal ME, Marits P, Thorn M, Ohlsson R, Winqvist O: FOXP3 promoter demethylation reveals the committed Treg population in humans. *PLoS One* 2008;3:e1612.
- Jenmalm MC, Björkstén B: Cord blood levels of immunoglobulin G subclass antibodies to food and inhalant allergens in relation to maternal atopy and the development of atopic disease during the first 8 years of life. *Clin Exp Allergy* 2000;**30**:34-40.
- 48 Sandberg M, Frykman A, Ernerudh J, Berg G, Matthiesen L, Ekerfelt C, Nilsson LJ, Jenmalm MC: Cord blood cytokines and chemokines and development of allergic disease. *Pediatr Allergy Immunol* 2009;**20**:519-527.
- Tsuda H, Michimata T, Hayakawa S, Tanebe K, Sakai M, Fujimura M, Matsushima K, Saito S: A Th2 chemokine, TARC, produced by trophoblasts and endometrial gland cells, regulates the infiltration of CCR4+ T lymphocytes into human decidua at early pregnancy. *Am J Reprod Immunol* 2002;**48**:1-8.
- Mjösberg J, Berg G, Jenmalm MC, Ernerudh J: FOXP3+ regulatory T cells and T helper 1, T helper 2, and T helper 17 cells in human early pregnancy decidua. *Biol Reprod* 2010;**82**:698-705.
- Piccinni MP, Beloni L, Livi C, Maggi E, Scarselli G, Romagnani S: Defective production of both leukemia inhibitory factor and type 2 T-helper cytokines by decidual T cells in unexplained recurrent abortions. *Nat Med* 1998;**4**:1020-1024.

- Liu CA, Wang CL, Chuang H, Ou CY, Hsu TY, Yang KD: Prenatal prediction of infant atopy by maternal but not paternal total IgE levels. *J Allergy Clin Immunol* 2003;**112**:899-904.
- Prescott SL, Holt PG, Jenmalm MC, Björkstén B: Effects of maternal allergenspecific IgG in cord blood on early postnatal development of allergen-specific Tcell immunity. *Allergy* 2000;**55**:470-475.
- 54 Sandberg M, Frykman A, Jonsson Y, Persson M, Ernerudh J, Berg G, Matthiesen L, Ekerfelt C, Jenmalm MC: Total and allergen-specific IgE levels during and after pregnancy in relation to maternal allergy. *J Reprod Immunol* 2009;81:82-88.
- Prescott SL, Breckler LA, Witt CS, Smith L, Dunstan JA, Christiansen FT: Allergic women show reduced T helper type 1 alloresponses to fetal human leucocyte antigen mismatch during pregnancy. *Clin Exp Immunol* 2010;**159**:65-72.
- Paul WE, Zhu J: How are Th2-type immune responses initiated and amplified? *Nat Rev Immunol* 2010;**10**:225-235.
- Zaghouani H, Hoeman CM, Adkins B: Neonatal immunity: faulty T-helpers and the shortcomings of dendritic cells. *Trends Immunol* 2009;**30**:585-591.
- Rose S, Lichtenheld M, Foote MR, Adkins B: Murine neonatal CD4+ cells are poised for rapid Th2 effector-like function. *J Immunol* 2007;**178**:2667-2678.
- Pfefferle PI, Buchele G, Blumer N, Roponen M, Ege MJ, Krauss-Etschmann S, Genuneit J, Hyvarinen A, Hirvonen MR, Lauener R, Pekkanen J, Riedler J, Dalphin JC, Brunekeef B, Braun-Fahrlander C, von Mutius E, Renz H: Cord blood cytokines are modulated by maternal farming activities and consumption of farm dairy products during pregnancy: the PASTURE Study. *J Allergy Clin Immunol* 2010;**125**:108-115 e101-103.
- Roduit C, Wohlgensinger J, Frei R, Bitter S, Bieli C, Loeliger S, Buchele G, Riedler J, Dalphin JC, Remes S, Roponen M, Pekkanen J, Kabesch M, Schaub B, von Mutius E, Braun-Fahrlander C, Lauener R: Prenatal animal contact and gene expression of innate immunity receptors at birth are associated with atopic dermatitis. *J Allergy Clin Immunol* 2011;**127**:179-185, 185 e171.
- Böttcher MF, Jenmalm MC, Björkstén B: Immune responses to birch in young children during their first 7 years of life. *Clin Exp Allergy* 2002;**32**:1690-1698.
- Hooper LV, Macpherson AJ: Immune adaptations that maintain homeostasis with the intestinal microbiota. *Nat Rev Immunol* 2010;**10**:159-169.
- 63 Sjögren YM, Tomicic S, Lundberg A, Böttcher MF, Björkstén B, Sverremark-Ekstrom E, Jenmalm MC: Influence of early gut microbiota on the maturation of childhood mucosal and systemic immune responses. *Clin Exp Allergy* 2009;**39**:1842-1851.
- Travis J: On the origin of the immune system. *Science* 2009;**324**:580-582.
- Lee YK, Mazmanian SK: Has the microbiota played a critical role in the evolution of the adaptive immune system? *Science* 2010;**330**:1768-1773.
- Björkstén B, Sepp E, Julge K, Voor T, Mikelsaar M: Allergy development and the intestinal microflora during the first year of life. *J Allergy Clin Immunol* 2001;**108**:516-520.
- Kalliomäki M, Kirjavainen P, Eerola E, Kero P, Salminen S, Isolauri E: Distinct patterns of neonatal gut microflora in infants in whom atopy was and was not developing. *J Allergy Clin Immunol* 2001;**107**:129-134.

- 68 Sjögren YM, Jenmalm MC, Böttcher MF, Björkstén B, Sverremark-Ekström E: Altered early infant gut microbiota in children developing allergy up to 5 years of age. *Clin Exp Allergy* 2009;**39**:518-526.
- Wang M, Karlsson C, Olsson C, Adlerberth I, Wold AE, Strachan DP, Martricardi PM, Åberg N, Perkin MR, Tripodi S, Coates AR, Hesselmar B, Saalman R, Molin G, Ahrné S: Reduced diversity in the early fecal microbiota of infants with atopic eczema. *J Allergy Clin Immunol* 2008;**121**:129-134.
- Forno E, Onderdonk AB, McCracken J, Litonjua AA, Laskey D, Delaney ML, Dubois AM, Gold DR, Ryan LM, Weiss ST, Celedon JC: Diversity of the gut microbiota and eczema in early life. *Clin Mol Allergy* 2008;**6**:11.
- Adlerberth I, Lindberg E, Aberg N, Hesselmar B, Saalman R, Strannegård IL, Wold AE: Reduced enterobacterial and increased staphylococcal colonization of the infantile bowel: an effect of hygienic lifestyle? *Pediatr Res* 2006;**59**:96-101.
- 72 Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, Knight R: Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci U S A* 2010;**107**:11971-11975.
- Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, Egholm M, Henrissat B, Heath AC, Knight R, Gordon JI: A core gut microbiome in obese and lean twins. *Nature* 2009;**457**:480-484.
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Jian M, Zhou Y, Li Y, Zhang X, Qin N, Yang H, Wang J, Brunak S, Dore J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J, Bork P, Ehrlich SD: A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 2010;464:59-65.