Photochemical mineralisation in a boreal brown water lake: considerable temporal variability and minor contribution to carbon dioxide production

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Abstract. Sunlight induces photochemical mineralisation of chromophoric dissolved organic matter (CDOM) to dissolved inorganic carbon (DIC) in inland waters, resulting in carbon dioxide (CO2) emissions to the atmosphere. Photochemical rate modelling is used to determine sunlight-induced CO2 emissions on large spatial and temporal scales. A sensitive model parameter is the wavelength-specific photochemical CDOM reactivity, the apparent quantum yield (AQY). However, the temporal variability of AQY spectra within inland waters remains poorly constrained. Here, we studied a boreal brown water lake in Sweden. We measured AQY spectra for photochemical DIC production monthly between June and November 2014 and parameterised a photochemical rate model. The total AQY between 280 and 600 nm increased about 3-fold during the open-water period, likely due to a high rainfall event with consecutive mixing in autumn that increased availability of highly photoreactive CDOM. However, the variability in AQY spectra over time was much smaller than previously reported variability in AQY spectra between lakes. Yet, using either the AQY spectrum from the least or from the most photoreactive water sample resulted in a 5-fold difference in simulated annual DIC photoproduction (2012–2014), with 2.0 ± 0.1 and 10.3 ± 0.7 g C m\(^{-2}\) yr\(^{-1}\), respectively. This corresponded to 1 and 8 % of the mean CO2 emissions measured from this lake. We conclude that (1) it may be recommendable to conduct repeated AQY measurements throughout the season for more accurate simulation of annual photochemical DIC production in lakes and (2), in agreement with previous studies, direct CDOM photomineralisation makes only a minor contribution to mean CO2 emissions from Swedish brown water lakes.

1 Introduction

Inland waters play a substantial role in carbon cycling (Cole et al., 2007; Battin et al., 2009; Tranvik et al., 2009). The major carbon fluxes occurring in inland waters are burial in sediments and mineralisation followed by carbon dioxide (CO2) emission into the atmosphere. A substantial fraction of the CO2 emissions is attributed to microbial mineralisation of dissolved organic carbon (DOC) (del Giorgio et al., 1997; Bertilsson and Tranvik, 2000). According to the first global upscaling study, up to about one-tenth of the CO2 emissions from lakes and reservoirs are directly sunlight induced (Koehler et al., 2014). However, the importance of sunlight for carbon processing varies strongly between systems and studies (Granéli et al., 1996; Bertilsson and Tranvik, 2000). Also, sunlight contributes to CO2 production via photochemical mineralisation of chromophoric dissolved organic matter (CDOM) (Granéli et al., 1996; Ziegler and Benner, 2000). According to the first global upscaling study, up to about one-tenth of the CO2 emissions from lakes and reservoirs are directly sunlight induced (Koehler et al., 2014). However, the importance of sunlight for carbon processing varies strongly between systems and studies (Granéli et al., 1996; Molot and Dillon, 1997; Ziegler and Benner, 2000; Cory et al., 2014).

Measuring photochemical DOC mineralisation, equivalent to photochemical production of dissolved inorganic carbon (DIC), in the field is challenging and seldom conducted (Salonen and Vähätalo, 1994; Granéli et al., 1996). Photochemical rate modelling is used to obtain DIC photoproduction
estimates at large spatial and temporal scales. Model parameterisation requires wavelength-specific irradiance, CDOM absorbance, attenuation and photochemical CDOM reactivity, i.e. the apparent quantum yield (AQY) defined as DIC produced per mol photons absorbed (Fichot and Miller, 2010; Koehler et al., 2014). The AQY is a sensitive model parameter but until now spectra have only been published from a small number of lakes (Vähätalo et al., 2000; Vähätalo and Wetzel, 2004; Koehler et al., 2014; Cory et al., 2014; Vachon et al., 2016), and temporal variability in AQY spectra within individual systems is even less studied (Cory et al., 2014; Vachon et al., 2016). Given the limited knowledge on spatial and temporal variability of AQY spectra the first large-scale modelling study of photochemical CDOM mineralisation in inland waters assumed that AQY spectra determined for single systems and on single occasions represented photochemical reactivity on larger spatial and temporal scales (Koehler et al., 2014).

However, temporal variability in AQY spectra is to be expected. For example, photochemical DIC production can increase with increasing CDOM aromatic content, increasing iron concentrations or decreasing pH (Gao and Zepp, 1998; Bertilsson and Tranvik, 2000; Anesio and Granéli, 2004; Stubbins et al., 2010). An important process that may influence CDOM quality on a seasonal scale is photobleaching, where CDOM is transformed to less coloured and less aromatic compounds (Brinkmann et al., 2003; Müller et al., 2014). Consequently, CDOM can become less photoreactive after light exposure (Lindell et al., 2000; Gonsior et al., 2013), and this “light dose dependence” may be especially important at high latitudes (Zhang et al., 2006). Temporal fluctuations may also be caused by photo-reactive terrestrial CDOM entering aquatic systems through heavy rainfall and runoff (Spencer et al., 2010; Hughes et al., 2013). For example, studies in a tropical systems observed the largest and smallest photochemical mineralisation rates during rainy and dry season, respectively (Amado et al., 2006; Suhett et al., 2007).

In this study, we examined temporal variability in photochemical reactivity and photochemical DIC production in a small brown water lake in Sweden. We then evaluated differences in photochemical DIC production simulated using a photochemical rate model with time-constant vs. repeatedly measured AQY spectra. Finally, we assessed the contribution of mean annual photochemical DIC production to total mean CO₂ emission from this lake.

2 Material and methods

2.1 Study lake and sampling

Erssjön (58°37’ N, 12°16’ E) is a small brown water lake (59 997 m², mean depth 1.3 m, maximum depth 4.4 m) in the Bäveån catchment in southwest Sweden (Fig. 1a). The lake is mostly surrounded by forest, mainly spruce and birch, and some agricultural land, and is part of the Skogaryd Research Site (Klemetchsson et al., 2010). In 2014, the ice disappeared from lake Erssjön on 25 February (S. Peter, personal communication, 2014) and the lake remained ice free until 31 December. For this study, 2 L of surface water was grab sampled into acid-washed polyethylene bottles in the middle of the lake, monthly between April and November 2014. The samples were kept dark and cold (<10 °C) until and during transport to Uppsala University within 1 to 3 days. Upon arrival, the water was filtered sequentially through pre-combusted glass fibre filters (Whatman GF/F, GE Healthcare, Little Chalfont, Buckinghamshire, UK) and 0.2 µm polyethersulfone membrane filters (Supor®-200, Pall Corporation, Ann Arbor, Michigan, USA) into glass bottles. Filtration through the 0.2 µm membrane filters, which was conducted to minimise microbial abundance and hence microbial respiration during the irradiation experiments (Sect. 2.3), reduced the integrated CDOM absorbance between 300 and 600 nm by 4.4 % compared to that of GF/F filtrate. The samples were wrapped in aluminium foil and kept at 4 °C until further analysis within 3 weeks.

2.2 Chemical and optical water properties

DOC concentrations were measured with a total carbon analyser (Shimadzu TOC-L, Shimadzu Corporation, Kyoto, Japan), as non-purgeable organic carbon (NPOC) concentration. UV–Vis absorbance spectra (200 to 600 nm) of filtered water were measured in a 1 cm quartz cuvette using a Lambda35 UV–Vis Spectrometer (PerkinElmer Lambda 25, Perkin Elmer, Waltham, USA). Based on the Beer–Lambert law, absorption coefficients a (m⁻¹) were calculated as

\[
\alpha = \frac{A \ln 10}{L}
\]
where $A$ is absorbance (dimensionless) and $L$ is optical path length (m) (Kirk, 2010). The specific UV absorption coefficient at 254 nm (SUVA$_{254}$; L mg C$^{-1}$ m$^{-1}$), a commonly used indicator of DOC aromaticity (Weishaar et al., 2003), was calculated as the ratio between $a_{254}$ and the DOC concentration (mg C L$^{-1}$). Synchronous fluorescence scans were obtained using a FluoroMax-4 Spectrofluorometer (FluoroMax-4, Jobin Yvon, Horiba, Kyoto, Japan), with excitation-emission matrices (EEMs) between excitation wavelengths 250 to 445 nm with 5 nm increments, and emission wavelengths 300 to 600 nm with 4 nm increments. The EEMs were blank subtracted using a sample of Milli-Q water run on the same day, corrected for instrument biases and inner filter effects and normalised to Raman units (Lawaetz and Stedmon, 2009; Kothawala et al., 2013). Three commonly used indices were calculated at fixed excitation/emission wavelength pairs or regions (Coble et al., 2014; Gabor et al., 2014). All fluorescence corrections and analyses were performed using the FDOMcorr toolbox for MATLAB (Murphy et al., 2010).

For total nitrogen (TN) analysis, all nitrogen species were oxidised to nitrate using potassium persulfate and sodium hydroxide at high pressure and temperature in an autoclave. TN was determined spectrophotometrically by subtracting a blank and absorbance at 275 nm from absorbance at 220 nm (PerkinElmer Lambda 40 UV–Vis spectrometer, Perkin Elmer, Norwalk, CT, USA). EDTA (disodium diydrogen-ethylenediaminetetraacetat) was used for the calibration curve (Rand et al., 1976). Total phosphorus (TP) was converted to orthophosphate using oxidative hydrolysis with potassium persulfate in acid solution at high pressure and temperature in an autoclave, and to phosphorus molybdate by reaction with ammonium molybdate, which was then reduced with ascorbic acid, accelerated by antinome. The samples were analysed spectrophotometrically at 882 nm as molybdate reactive phosphorus (PerkinElmer Lambda 40) (Menzel and Corwin, 1965; Murphy and Riley, 1958). TP concentrations measured for the LAGGE project were used (M. Wallin, unpublished data).

### 2.3 Apparent quantum yield

The wavelength-specific CDOM reactivity towards photochemical DIC production, i.e. the AQY defined as mol DIC produced per mol CDOM absorbed photons, was determined monthly between June and November 2014 similarly as described in Koehler et al. (2014). The measurements from April and May could not be used due to failure of the DIC analyser. Specifically, to minimise initial DIC concentration, the samples were acidified (10 % HCl to pH$<3$), bubbled with nitrogen gas for 25 min to remove the CO$_2$, and re-adjusted to the original pH using 1M NaOH. The amount of HCl and NaOH added never exceeded 0.5 % of the sample volume. The water was re-filtered with 0.2 µm Supor$^{	ext{®}}$-200 filters to minimise bacterial abundance and hence respiration during subsequent irradiation. During this filtration step the water, in which oxygen concentrations were reduced during bubbling with N$_2$, was also aerated again. The water was then filled into cylindrical glass vials with flat quartz top (50 mL volume; Fig. S1). The incubation vials were soaked in 10 % HNO$_3$ for at least 10 hours and rinsed thoroughly with Milli-Q water before and after each experiment. To systematically manipulate the irradiance field, cut-off filters (CVI Laser Corporation, obtained from former Gamma Optronik AB, Sweden and Oriel Instruments, Newport Corporation, Irvine, California) that cut-off irradiance with wavelengths below 455, 420, 380, 350, 320, 309 or 280 nm (Fig. S1) were placed on top of the vials. All filters and dark controls, where a black lid was attached to the vial, were used in triplicate. Thin needles were inserted through the septa covering one of the vial outlets to enable pressure release during irradiation in the solar simulator. Using three vials with and three vials without a needle through the septum, which were filled with a standard of 1500 ppb IC and left at room temperature for 24 h, we verified that this did not affect DIC concentration in the vessel ($p = 0.113$). Then, the samples were irradiated for 5 hours using a solar simulator (Q-Sun 1000 Xenon test chamber, Q-panel Lab Products Europe, Bolton, UK) set to 0.59 W m$^{-2}$ at 340 nm (calibrated with the instrument’s CR20 Calibration Radiometer). During irradiation, the samples were standing in a cooled water bath, maintaining the temperature around the vials at approximately 25°C. Initial and final DIC concentrations were measured from each vial with the Shimadzu TOC-L analyser, and the photochemical DIC production in each vial was calculated as the difference between the final and initial DIC concentration, minus the mean production in the dark controls. A calibration curve was created before each run, using the auto-dilution function to create six standards of different concentrations from a 5 or 10 ppm solution that was freshly prepared from a 1000 ppm IC stock solution ($R^2 > 0.998$) (Shimadzu user manual). DIC concentrations were measured in a minimum of five injections of 150 µL, resulting in SD (standard deviation)$<0.5$ ppb and/or CV (coefficient of variation) $<2\%$. In the June experiment the “dark DIC production” was $-0.2$ to $-17$ ppb. We suspect this was due to a slight offset in the calibration of the instrument during the measurements and/or difficulty to detect very low DIC concentrations, and set the control values to zero. In the August experiment, the dark production of one control set was considerably higher than usual. Since the acid-washing step had been missed during cleaning of these three vessels, we suspect the high concentrations were caused by contamination. Therefore, the values of the other control set were used for calculating photochemical DIC production. This affected the resulting AQY spectrum only to a minor extent (Fig. S2b in the Supplement). Across experiments, DIC production in the dark controls averaged 26.2± 4.6 ppb, corresponding to 3 and 24 % of the average DIC production under the 250 and 455 nm cut-off filter, respectively. On eight occasions throughout the study period, irradiance spec-
tra (280–600 nm) were measured at the location of each vial using a spectrometer (BLACK Comet UV–Vis, StellarNet Inc., Tampa, Florida, USA) equipped with a fibre optic cable (STEF600-UV–Vis-SR, StellarNet) and a cosine receptor for UV–Vis near-infrared irradiance (STE-CR2, StellarNet). Absorbed photons were calculated accounting for the inner filter effect (Hu et al., 2002). The calculated number of CDOM-absorbed photons was in good agreement with CDOM-absorbed photons determined using nitrite ultraviolet–actinometry, where the photon exposure of an irradiated sample is quantified from the photochemical production of salicylic acid formed during reaction of the hydroxide radical with benzoic acid (Jankowski et al., 1999, 2000). The response bandwidth was verified, and the photoproduced salicylic acid was detected using fluorescence spectrophotometry (SPEX FluoroMax-4; Jankowski et al., 1999). CDOM-absorbed photons determined with the spectrally resolved calculation used during AQY determination and the broadband actinometry differed by a factor of 1.43 ± 0.04 under the complete irradiance spectrum in the solar simulator.

AQY spectra were calculated using weighted parameter optimisation (Rundel, 1983) to an exponential function

\[ \Phi = e^{-\left(m_1 + m_2 (\lambda - 290)\right)} \]

where \( \Phi \) is the AQY of DIC photoproduction (mol DIC mol photons\(^{-1}\)), \( \lambda \) is the wavelength (nm) and \( m_1 \) and \( m_2 \) are fit parameters (Johannessen and Miller, 2001), using the Nelder Mead simplex minimisation algorithm (Nelder and Mead, 1965) implemented in the function optim in R 3.1.0 (R Core Team, 2014), and using a set of different starting values to verify stability of the solution. The total AQY (AQY\(_{\text{total}}\)) was calculated using the DIC production measured under full irradiance (280 nm filter) divided by CDOM-absorbed photons integrated from 280 to 600 nm. For uncertainty estimation we used bootstrapping (Ritz and Streibig, 2008; Crawley et al., 2012), where we resampled the monthly measured photochemical DIC production with replacement, i.e. duplicates are possible within each bootstrap resample (6000 times), assigned the respective CDOM-absorbed photons, and fitted AQY spectra to each bootstrap data set. We give the 2.5 and 97.5 % quantiles of the resulting bootstrap distribution of parameter estimates as 95 % confidence intervals. Kernel density estimation was used to estimate the probability density function for the bootstrap distributions of parameter estimates. To obtain simultaneous pointwise confidence intervals (Fig. 2) we used the 6000 bootstrap parameter estimates to predict the AQY at five discrete wavelengths, about midway between the cut-off filters used during the irradiation experiments (295, 330, 365, 400 and 435 nm). The confidence level was Bonferroni corrected to reduce the family-wise type I error rate according to \( (1 - \frac{\alpha}{n}) \cdot 100\% \), where \( \alpha \) is the significance level and \( n \) is the number of simultaneous calculations.

To statistically test the temporal variability in AQY we calculated the difference in the discrete AQY values calculated above between adjacent sampling months (comparing June to July, July to August, etc., including November to June. Again, the confidence level was adjusted for multiple testing using the Bonferroni correction. A temporal difference \( (p \text{ value} \leq 0.05) \) exists when the obtained 95 % confidence intervals of the differences between adjacent months exclude zero in at least one case.

### 2.4 In situ photochemical DIC production

During 23 to 25 July 2014, we determined in situ photochemical DIC production rates similarly as described in Granéli et al., 1996. Specifically, we filled filtered lake water (0.2 µm membrane filters) into quartz tubes (38 mL, 2 cm diameter) and corresponding borosilicate dark control tubes wrapped in aluminium foil. Three quartz and two to three dark tubes were attached horizontally to steel wire racks, which were secured to a floating wooden frame that was kept in place with two anchors. This set-up was duplicated and the two frames were positioned in the lake at least 50 m from the shoreline (Fig. 1b; red dots). The racks with the tubes were positioned such that the centre of the tubes was positioned at 1, 4 and 8 cm water depth and well within the frame, so that no shading occurred (Fig. 1c). During the 2-day incubation period the anchors sank into the sediment and pulled the frames down by approximately 1 cm. After incubation, all
tubes were wrapped in aluminium foil, placed with cooling blocks in cooling boxes for transport, and stored at 4 °C until analysis at Uppsala University within 2 days. Initial DIC concentration was measured from one water sample taken and filtered at the start of the incubation as described above, and kept cold and dark until analysis after 3 days. Final DIC concentrations were measured directly from the incubation tubes and averaged for the three pseudo-replicate tubes. In one case the measured value of one of the dark triplicates was about 35% higher than all other dark values. This sample was considered to be contaminated and excluded from the calculations. The DIC production at the different water depths was then calculated as the mean of the two set-ups and standardised to mg C m⁻³ d⁻¹.

2.5 Photochemical rate modelling

Using photochemical rate modelling (Eq. 3), DIC photoproduction was simulated for the open-water periods of 2012 to 2014 as

$$
\Psi_{DIC}^{day} = \int_{\lambda_{min}}^{\lambda_{max}} E_{od}^{day}(\lambda, 0^-) a_\lambda(\lambda) e^{-(K_d(\lambda)z)} \phi(\lambda) d\lambda. \tag{3}
$$

The model calculates the daily photochemical DIC production rate (\(\Psi_{DIC}^{day}, \text{mol} \ C \ m^{-3} \ d^{-1} \ nm^{-1}\)) over water depth (\(z, \text{m}\)) based on daily-integrated downwelling scalar irradiation just below the water surface (\(E_{od}^{day}(\lambda, 0^-), \text{mol} \ photons \ m^{-2} \ d^{-1} \ nm^{-1}\)), CDOM absorption coefficient (\(a_\lambda, \text{m}^{-1}\)), vertical attenuation coefficient for downwelling irradiance (\(K_d, \text{m}^{-1}\)) and the apparent quantum yield (\(\phi(\lambda), \text{mol} \ DIC \ mol \ photons^{-1}\)) over the photochemically relevant wavelength range (\(\lambda, 280-600 \text{nm}\)) (Fichot et al., 2010; Koehler et al., 2014). Daily-integrated clear-sky irradiance spectra were obtained using the libRadtran model (version 1.6) for radiative transfer (Mayer et al., 2005), parameterised and cloud corrected as described in Koehler et al. (2014). For the year 2014, for which monthly AQY spectra were measured between June and November, we used four different AQY parameterisations and assessed their influence on the simulated photochemical DIC production. In the first parameterisation, we assumed that the measured AQY and absorbance spectra were representative for 1 month around the sampling dates. The spectra measured in June were also used for the open-water period prior to June, and the spectra measured in November were used until the end of the open-water period in December. In the second parameterisation, we assumed that the AQY spectrum fitted through all data points obtained between June and November is a representative description of the photochemical reactivity in the lake. The absorbance spectra were again used for 1 month around the sampling dates. In the third and fourth parameterisation, we assumed that the observed most and least photosynthetic water samples were representative throughout the whole open-water period, respectively.

SUVA\(_{254}\) was calculated for the years 2012 to 2014, using data from this study as well as absorbance spectra and TOC concentrations measured in 2012 and 2013 (M. Wallin, unpublished data). Since no actual ice-on and ice-off dates were available for lake Erssjön in 2012 and 2013, the long-term average (1970–2007) ice-cover dates for the nearby (19 km) lake Ellenösjön were used (3 April to 7 December; SMHI, 2013).

We also compared simulated photochemical DIC production with the in situ measured rates. In order to compare with the rates measured in the incubation tubes, we integrated the simulated sunlight-induced DIC production rates over the respective depth intervals and for the same time period as the in situ measurement. Since the duration of the incubation was only 2 days, hourly rather than daily irradiance spectra were used. We assumed that the quartz tubes did not interfere with irradiance. While, in reality, the quartz tubes will affect the number and optical path length of the photons entering the tube, we considered this effect minor compared to other uncertainties during the in situ measurements (see Discussion). The absorbance coefficients and apparent quantum yield were obtained from water sampled on the last day of the incubation (the July sample of this study; Fig. 1).

2.6 Total CO\(_2\) emissions

Total CO\(_2\) emissions from the lake surface were measured using plastic floating chambers of volume 6.3 L and area 0.07 m\(^2\), which were covered with aluminum tape to reflect sunlight thereby minimising internal heating, equipped with Styrofoam collars to enable floating and anchored to the lake bottom. The chamber walls extended 3 cm into the water on deployment. Mini CO\(_2\) sensors (CO\(_2\) Engine® ELG, SenseAir AB, Sweden; measuring range 0–10 000 ppm) were fitted inside the chamber and programmed to log CO\(_2\) concentrations every 5 min (Bastviken et al., 2015). Three chambers were deployed over water depths of 0.5, 2.5 and 4 m (Fig. 1b; white dots). Before flux measurements, the chambers were vented using a 20 cm long PVC tube fitted with a three-way luer-lock stopcock (Becton-Dickinson, USA). After venting, the chambers were closed for 30 min, and the rate of change in CO\(_2\) concentration inside the chamber was calculated using linear regression. When the change of CO\(_2\) concentrations over time was non-linear, with \(R^2<0.9\), we discarded the time series. The rates were converted to moles using the ideal gas law and divided by area and time to obtain emissions. Measurements were made approximately every 2 weeks during June to October 2012 and April to November 2013. During each visit, emissions were measured on 2 consecutive days.

2.7 Statistical analyses

Two sample \(t\) tests were used to test for differences between DIC production under the cut-off filters and the dark con-
to almost 5% of the annual rainfall in 2014, and another 97 mm precipitation was observed during the remainder of the month (SMHI, 2015). Subsequently, from August until November, DOC concentrations and absorbance coefficients were approximately 50% higher than earlier in the year (\( p_{\text{DOC}} = 0.022 \), \( p_{\text{SUVA}_{254}} = 0.009 \), \( p_{\text{a}_{250}} = 0.025 \)), while \( p \) and \( \text{SUVA}_{254} \) remained similar. TN and TP were similar across the open-water period with the exception of August, when TN was approximately twice as high. The fluorescence index (FI) increased slightly throughout the study period (\( p = 0.003 \)), whereas the freshness index (\( \beta : \alpha \)) showed no apparent pattern over time. The humification index (HIX) decreased in spring and early summer, increased towards autumn and then decreased again (Table 1). DOC concentrations, \( a_{250} \) and \( \text{SUVA}_{254} \) were similar during 2012–2014 (Table 2).

### 3 Results

#### 3.1 Chemical and optical water properties

Water chemical and optical properties were similar in lake Erssjön from April to July 2014 (Table 1; Fig. S2a). Sampling in August was preceded by a period of high rainfall with 46 mm precipitation within 7 days. This corresponded to almost 5% of the annual rainfall in 2014, and another 97 mm precipitation was observed during the remainder of the month (SMHI, 2015). Subsequently, from August until November, DOC concentrations and absorbance coefficients were approximately 50% higher than earlier in the year (\( p_{\text{DOC}} = 0.022 \), \( p_{\text{SUVA}_{254}} = 0.009 \), \( p_{\text{a}_{250}} = 0.025 \)), while \( p \) and \( \text{SUVA}_{254} \) remained similar. TN and TP were similar across the open-water period with the exception of August, when TN was approximately twice as high. The fluorescence index (FI) increased slightly throughout the study period (\( p = 0.003 \)), whereas the freshness index (\( \beta : \alpha \)) showed no apparent pattern over time. The humification index (HIX) decreased in spring and early summer, increased towards autumn and then decreased again (Table 1). DOC concentrations, \( a_{250} \) and \( \text{SUVA}_{254} \) were similar during 2012–2014 (Table 2).

#### 3.2 Apparent quantum yield

The DIC production under full irradiance (\( p = 0.002 \)) and the AQY (\( p = 0.008 \)) increased throughout the sampling year, while there was no significant change in CDOM-absorbed photons (Table 3). The monthly AQY spectra, evaluated at five discrete wavelengths and tested simultaneously,
Table 3. Mean (±SE) photochemical DIC production under the full irradiance spectrum in the solar simulator, and absorbed photons as well as the total AQY in the wavelength range 280–600 nm; parameter estimates for the fitted AQY spectra (Eq. 2), information on performance to reproduce the observations ($R^2$, regression slope and nRMSE) and areal photochemical DIC production in 2014 using the respective AQY spectra. Values in parentheses give diagnostics and simulation results when single AQY spectra were used to predict photochemical DIC production observed during all six irradiation experiments.

<table>
<thead>
<tr>
<th>AQY</th>
<th>DIC production under full irradiance (μmol L$^{-1}$ h$^{-1}$)</th>
<th>CDOM-absorbed photons$_{280–600}$ (mol m$^{-2}$ h$^{-1}$)</th>
<th>AQY$_{total}$ (mmol DIC mol photons$^{-1}$)</th>
<th>$m_1$</th>
<th>$m_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>June</td>
<td>9.28 ± 0.72</td>
<td>3.12 ± 0.23</td>
<td>0.138 ± 0.003</td>
<td>5.776±0.518</td>
<td>0.003±0.007</td>
</tr>
<tr>
<td>July</td>
<td>7.54 ± 0.42</td>
<td>3.77 ± 0.26</td>
<td>0.093 ± 0.006</td>
<td>5.985±0.373</td>
<td>0.006±0.004</td>
</tr>
<tr>
<td>August</td>
<td>17.57 ± 0.90</td>
<td>3.97 ± 1.81</td>
<td>0.206 ± 0.007</td>
<td>5.846±0.454</td>
<td>0.002±0.007</td>
</tr>
<tr>
<td>September</td>
<td>19.90 ± 1.26</td>
<td>4.52 ± 0.31</td>
<td>0.204 ± 0.004</td>
<td>5.839±0.137</td>
<td>0.001±0.007</td>
</tr>
<tr>
<td>October</td>
<td>29.41 ± 1.76</td>
<td>4.02 ± 0.28</td>
<td>0.341 ± 0.016</td>
<td>5.782±0.316</td>
<td>0.003±0.003</td>
</tr>
<tr>
<td>November</td>
<td>33.87 ± 0.98</td>
<td>4.21 ± 0.24</td>
<td>0.375 ± 0.014</td>
<td>5.967±0.218</td>
<td>0.002±0.002</td>
</tr>
<tr>
<td>Monthly measured</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Pooled</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>6.350±0.672</td>
<td>0.006±0.005</td>
</tr>
</tbody>
</table>


$R^2$: coefficient of determination. nRMSE: normalised root mean squared error between observed and predicted DIC production. DIC$_{areal}$: areal DIC photo production rate simulated for the open-water season of 2014, 310 days between the ice-off and ice-on dates.

The in situ photochemical DIC production rates decreased sharply by about a factor of 5 from just below the water surface to 4 cm water depth (Fig. 3, black numbers). At 8 cm depth, DIC production did not differ between the irradiated and the dark tubes; i.e. the photochemical DIC production was below the detection limit. The simulated photochemical DIC production also decreased sharply with increasing water depth (Fig. 3, red curve). When assuming that the experimental tubes remained at the intended depth of incubation, the simulated photochemical DIC production differed by 114 % at 1 cm and by 22 % at 4 cm from the observed rates, and, in accordance with the measurements, was small at 8 cm depth (Fig. 3, red numbers). However, the racks to which the tubes were attached were pulled down by approximately 1 cm over the course of the 2-day incubation period as their anchors sank into the sediment. If the depth intervals of integration are taken to be 1 cm lower than the intended depths, the simulated DIC production rates differed by 38 % at 2 cm and by 9 % at 5 cm from the observed values (Fig. 3; red numbers in parentheses).

### 3.3 Observed vs. simulated photochemical DIC production rates

The in situ photochemical DIC production rates decreased sharply by about a factor of 5 from just below the water surface to 4 cm water depth (Fig. 3, black numbers). At 8 cm depth, DIC production did not differ between the irradiated and the dark tubes; i.e. the photochemical DIC production
Photochemical DIC production for the years 2012 and 2013, in combination with the measured absorbance spectra and simulated irradiance. Interannual variability in irradiance was very small (Fig. 4a) and hence, in combination with similar CDOM absorbance and the assumption that photoreactivity was similar as in 2014, simulated DIC photoproduction was similar across the years (Table 2; Fig. 4b). Simulating irradiance over the years 2004 to 2014 showed that the irradiance that lake Erssjön received in 2012 to 2014 was representative for the decadal mean (4.10×10^5±0.15×10^5 Wh m^{-2} yr^{-1}). During simulations we assumed that irradiance was not transmitted into the water column during the ice-covered period. If we instead would assume that the ice fully transmits irradiance or ice cover was absent, the yearly simulated photochemical DIC production would increase by 11 to 14%.

4 Discussion

The AQY spectra for photochemical DIC production, measured monthly between June and November 2014 in a boreal brown water lake, showed considerable seasonal variability, with the slope of the spectrum decreasing over the open-water season. Photochemical DIC production, simulated using photochemical rate modelling, made a minor contribution to the total CO₂ emissions observed from the same lake (Fig. 5). Hence, similar results from earlier studies in boreal Sweden (Jonsson et al., 2001; Koehler et al., 2014; Chmiel et al., 2016) were corroborated when considering temporal variability in photochemical reactivity as well as in total lake CO₂ emissions. Moreover, the good match between photochemical DIC production observed in situ and simulated rates (Fig. 2) supported that photochemical rate modelling is a suitable approach to investigate photochemical CDOM mineralisation in lakes and its contribution to carbon cycling on broader temporal and spatial scales. This highlights the potential to use a similar method for studying this process also in other climate zones, e.g. for tropical lakes, where the role of photochemical mineralisation for lake carbon cycling remains even less constrained than in boreal and temperate systems.

The DIC photoproduction rates observed in situ in the studied boreal brown water lake (Fig. 3) were comparable to rates in a Norwegian dystrophic lake (100 and 40 mg C m⁻³ d⁻¹ at 1 and 10 cm depth, respectively; Salonen and Vähätalo, 1994), five Swedish lakes (100–300 mg C m⁻³ d⁻¹ at 1 cm depth; Granéli et al., 1996) and in a Finnish humic lake (300 and 180 mg C m⁻³ d⁻¹ at 1 and 2.5 cm depth, respectively; Vähätalo et al., 2000). However, it is difficult to accurately measure DIC photoproduction rates in situ. Wind and wave action make it hard to exactly measure, adjust and stabilise the tubes at the intended depths of incubation. This is especially relevant in the case of a brown water lake like Erssjön, where DOC photomineralisation is confined to the upper centimetres of the water column and photochemical rates decrease rapidly with increa-

Figure 3. Photochemical DIC production rates observed in situ (±SE; black numbers) and simulated using photochemical rate modelling (red curve and average values over the intended depths of the experimental tubes). The frame to which the tubes were attached sank into the sediment by about 1 cm during the 2 incubation days. Simulated values adjusted to this change in incubation depths are given in parentheses.
The wavelength-specific photochemical reactivity is a critical and sensitive parameter in photochemical rate modelling (Fichot and Miller, 2010; Koehler et al., 2014; Cory et al., 2014). Nevertheless, the simulated and observed DIC photoproduction rates were similar (Fig. 3), giving confidence in the model parameterisation. Given the experimental difficulties, photochemical rate modelling is an attractive method for estimating photochemical DOC mineralisation, especially on large temporal and spatial scales.

The wavelength-specific photochemical reactivity is a critical and sensitive parameter in photochemical rate modelling (Fichot and Miller, 2010; Koehler et al., 2014; Cory et al., 2014). However, knowledge on its variability remains scarce. So far, AQY spectra for photochemical DIC production have only been reported for a small number of Arctic, boreal and temperate lakes (Vähätalo et al., 2000; Vähätalo and Wetzel, 2004; Koehler et al., 2014; Cory et al., 2014; Vachon et al., 2016). Information about temporal variability in AQY spectra across seasons within single lakes is even more rare, with only two studies so far where lake AQY spectra were repeatedly determined during the open-water season (Cory et al., 2014; Vachon et al., 2016). In this study, the AQY spectra determined monthly in a boreal brown water lake showed a decrease in slope (fit parameter $m_2$, Eq. 2) from June to November (Table 3; Fig. S2b, d in the Supplement). This suggests that the longer wavelengths contributed more to DIC photoproduction later in the season. However, the variability in AQY spectra over time ($\text{CV} = 0.11$ at $\lambda_{300}$) was much smaller (Fig. 1b) than the variability in AQY spectra between lakes of differing CDOM quality and quantity reported so far ($\text{CV} = 0.52$ at $\lambda_{300}$; Vähätalo et al., 2000; Vähätalo and Wetzel, 2004; Koehler et al., 2014; Vachon et al., 2016; AQY$_{300}$ of Toolik Lake from 29 June 2012, R. Cory, personal communication, 2014). Yet, given the high sensitivity of simulated DIC photoproduction towards both magnitude and slope of the AQY spectrum, applying AQY spectra measured at different times to the whole open-water period of 2014 resulted in up to 5.6-fold differences in simulated annual DIC photoproduction. Hence, depending on scale and scope of the study as well as feasibility, it may be recommendable to conduct repeated measurements of AQY spectra throughout the season for more accurate simulation of annual photochemical DIC production in lakes, as recently conducted in studies in the Arctic (Cory et al., 2014) and northern temperate and boreal Canada (Vachon et al., 2016).

While photobleaching is a relevant process regulating CDOM absorption on a seasonal scale in some humic boreal lakes (Müller et al., 2014), we did not observe net photochemical bleaching with a potentially associated reduction in DOM photoreactivity (Lindell et al., 2000). However, AQY spectra were only determined from June onwards, leaving the spring, in which photoreactivity may be high (Gonsior et al., 2013; Vachon et al., 2016) and bleaching most prevalent (Lindell et al., 2000; Zhang et al., 2006; Gonsior et al., 2013), unstudied. Values for the fluorescence index were around 1.3 throughout the season, indicating that the fluorescent DOM was mostly of terrestrial origin. Also the freshness index was stable, suggesting no major temporal changes in the proportion of recently produced fluorescent DOM from microbial origin (Gabor et al., 2014). A marked increase in DOC concentrations and absorbance in autumn (Table 1; Fig. S2a in the Supplement) was preceded by a high rainfall event (SMHI, 2015) and consecutive mixing of the lake (S. Peter, personal communication, 2014). Consistent

![Figure 4](https://example.com/fig4.png)

**Figure 4.** (a) Daily irradiance integrated over the wavelength range 280–600 nm. (b) Daily photochemical DIC production rate from 2012 to 2014 using the AQY spectrum with highest (November; primary y axis) and the lowest productivity (July; secondary y axis) measured in 2014. The grey shaded areas mark the ice-covered periods of the lake, during which we set DIC photoproduction to zero assuming no irradiance transmission (Petrov et al., 2005).
with the observed simultaneous increase in the humification index (Table 1), this event likely added a substantial amount of humified material to the lake, both from land and from the bottom water of the lake itself (Spencer, 2010; Gonsior et al., 2013; Hughes et al., 2013). Hence, rainfall events, mixing of the lake and potentially a shorter residence time towards autumn may have added fresh and more photoreactive material to the lake. Possibly, this masked photobleaching while increasing photoreactivity (Fig. 2). Similarly, rainfall and input of fresh terrestrial material increased CDOM photoreactivity in tropical lakes (Amado et al., 2006; Suhett et al., 2007). For tropical systems, which receive an even dose of sunlight throughout the year, the importance of photochemical reactivity in regulating temporal variability in photochemical DIC production may be expected to be higher than in boreal lakes, where temporal changes in photochemical reactivity interact with the pronounced seasonality in irradiance. Accordingly, CDOM photoreactivity and irradiance explained a similar amount of variability in photochemical mineralisation across seasons for three boreal and northern temperate lakes (Vachon et al., 2016).

Considering that photoreactions are constrained to a shallow top layer of the lake, the relative contribution of photochemistry to overall dynamics of DOC is uncertain. To address this, we compared the DIC photoproduction with the total CO₂ emissions that were measured from the lake. Assuming that all photoproduced DIC was emitted as CO₂ to the atmosphere, the mean simulated DIC photoproduction (7.9 ± 0.3–41.3 ± 2.9 mg C m⁻² d⁻¹; 2012–2014) contributes 1–8 % to the mean observed CO₂ emissions of 562.2 mg C m⁻² d⁻¹ (Fig. 5). Hence, the results of this detailed study in one Swedish brown water lake are in agreement with a large-scale modelling study for 1086 Swedish lakes, in which the contribution of mean annual DIC photoproduction to CO₂ emissions was about 12 % (Koehler et al., 2014). Also in agreement, direct photo-oxidation contributed about 7 % to the total DOC mineralisation in a large humic lake in northern Sweden (Jonsson et al., 2001), and 6 % in a small brown water lake in central Sweden (Chmiel et al., 2016). In a study based on 21 463 observations from lakes across Sweden, CO₂ emission ranged from 31.9 to 88.3 g C m⁻² yr⁻¹ (Humborg et al., 2010). Comparing our low and high estimate of simulated DIC photoproduction to these numbers would suggest a directly sunlight-induced contribution of 2 to 6 and 12 to 32 % to the total CO₂ emission, respectively. Besides the here studied direct effect of sunlight on DOC mineralisation, sunlight can also stimulate bacterial respiration by partially photo-oxidising DOC. The magnitude of this indirect effect can be as large as that of the direct effect (Lindell et al., 1995; Molot and Dillon, 1997; Bertilsson and Tranvik, 1998; Cory et al., 2014), resulting roughly in a doubling of the estimates presented here. We conclude that the contribution of sunlight to the CO₂ emissions from the studied Swedish brown water lake was small. This was also the case when taking temporal variability of

Figure 5. Box-and-whiskers plots of total measured CO₂ emissions, and minimum and maximum simulated photochemical DIC production, showing the median and first and third quartiles with the whiskers set at ±1.5 times the interquartile range and data outside this range given as circles. Note the log scale on the y axis.

AQY spectra into account. Even when using the AQY spectrum from the most photoreactive water sample for annual simulation and considering photo-stimulation of DOC mineralisation, the contribution of DOC phototransformations to the in-lake carbon cycling would still be minor.

5 Data availability

Supporting research data can be accessed at doi:10.13140/RG.2.1.5127.8329. Fluorescence and total CO₂ flux data and are available upon request from the corresponding author.

The Supplement related to this article is available online at doi:10.5194/bg-13-3931-2016-supplement.

Author contributions. B. Koehler designed the study. M. Groeneveld conducted laboratory and field experiments assisted by B. Koehler and L. Tranvik. M. Groeneveld and B. Koehler conducted photochemical rate modelling and data analysis. S. Natchimuthu conducted the total CO₂ flux measurements and analysed the flux data. M. Groeneveld wrote the manuscript with contributions and revision by B. Koehler, L. Tranvik and S. Natchimuthu.
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